



Università degli Studi di Bari Aldo Moro

Anno Accademico 2019/20

**Scuola di Specializzazione in
“Fisiopatologia della Riproduzione degli
Animali Domestici”**

Embryo quality assessment

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Summary (1)

- Historical aspects
- Physiology of embryo development in mammals
- Embryo metabolism
- Embryo genome activation (EGA)
- In vivo embryo culture systems
- In vitro embryo culture systems

Historical aspects

- Sreenan, 1968; Sreenan e Scanlon, 1968
(first reports of bovine **in vitro** embryo culture up to the blastocyst stage)
- Adams, 1968
(first report of bovine **in vivo** embryo culture in **rabbit oviducts** up to the hatching blastocyst stage)
- Gandolfi, 1986; Gandolfi and Moor, 1987
(first reports of bovine **in vitro** embryo culture up to the blastocyst stage on **oviductal epithelial cell monolayers**)

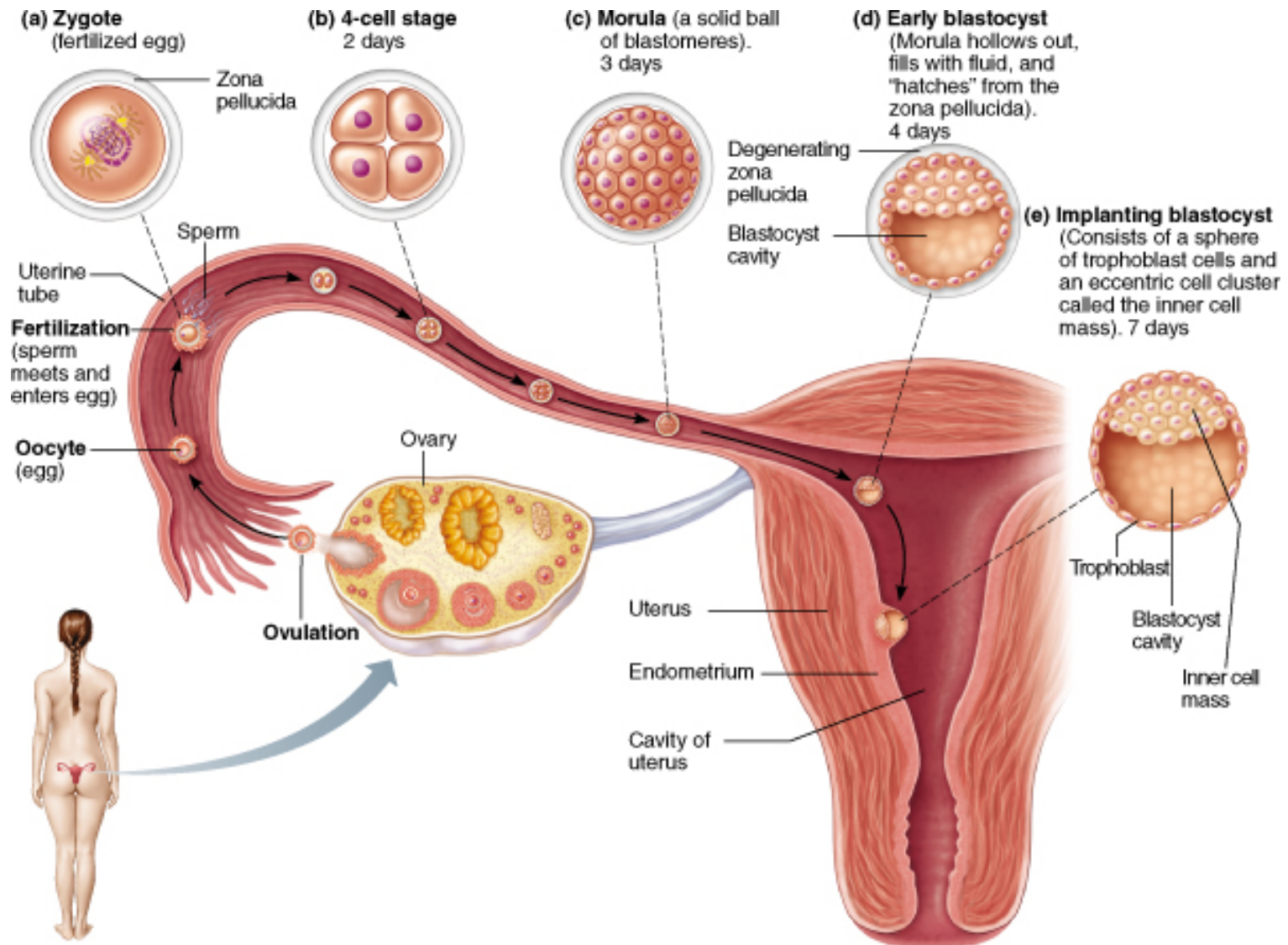


Studies on embryo/oviduct biochemical interactions

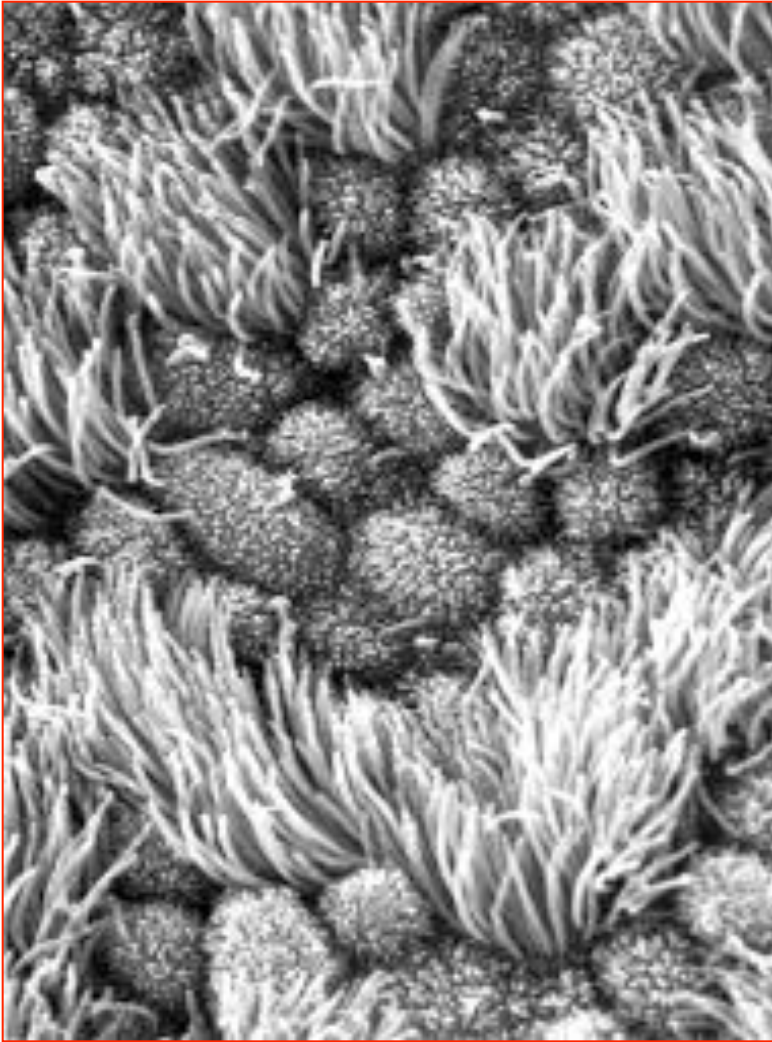
Physiology of embryo development in the pre-implantation period

- Oviductal microenvironment
- Stages of embryo development
- Cleavages
- Morula formation and compaction
- Blastocyst formation
- Embryo mortality

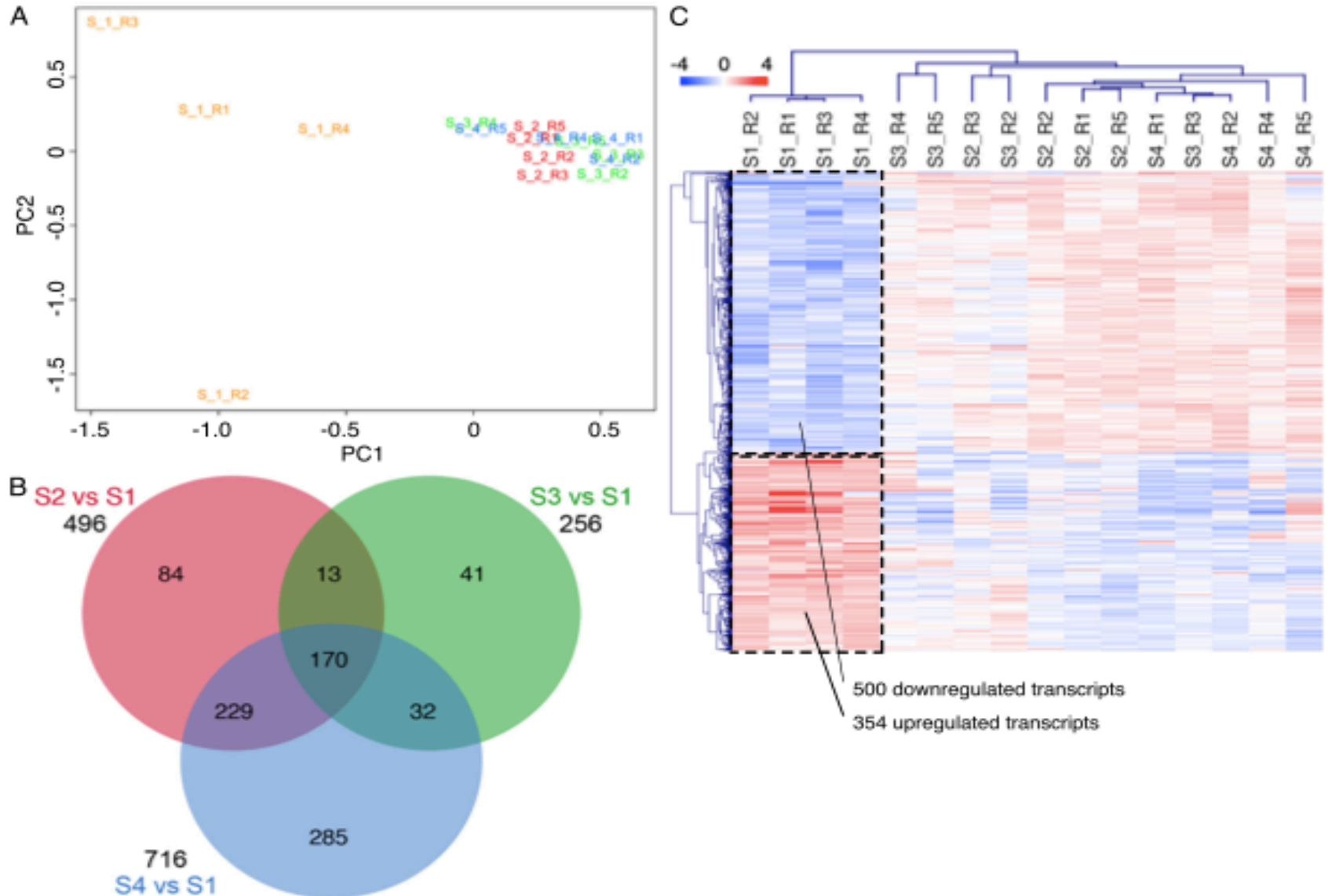
Stages of embryo development within the oviduct



The oviductal environment



Recently deciphered oviductal vesicles content in proteins, mRNA and small ncRNA across the estrous cycle in the cow (**13.197 genes**)



Stages of human embryo development



2PB 2PN



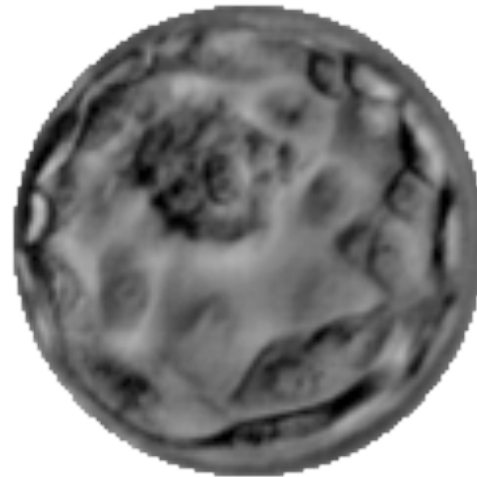
4 cell stage



8 cell stage

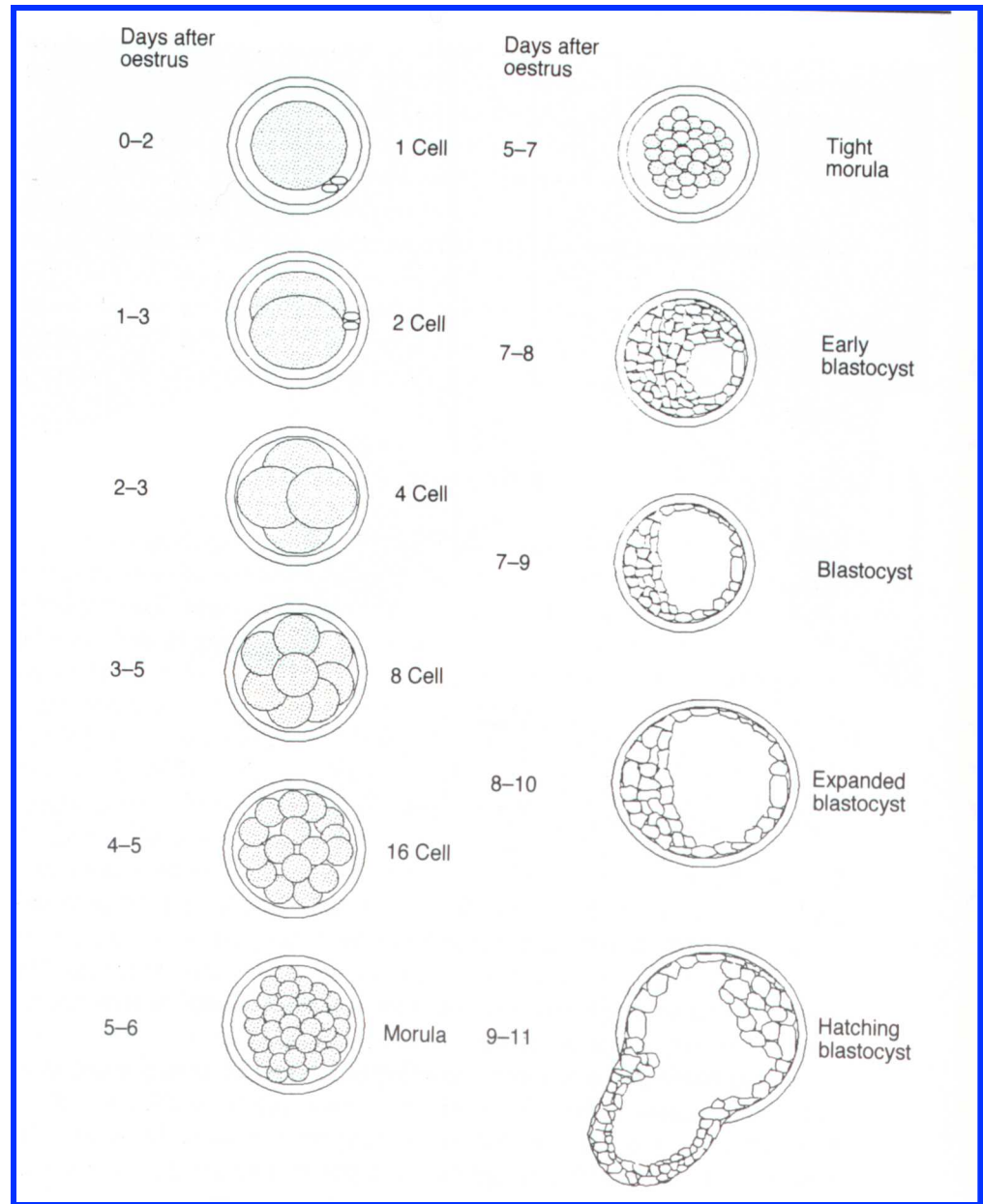


morula

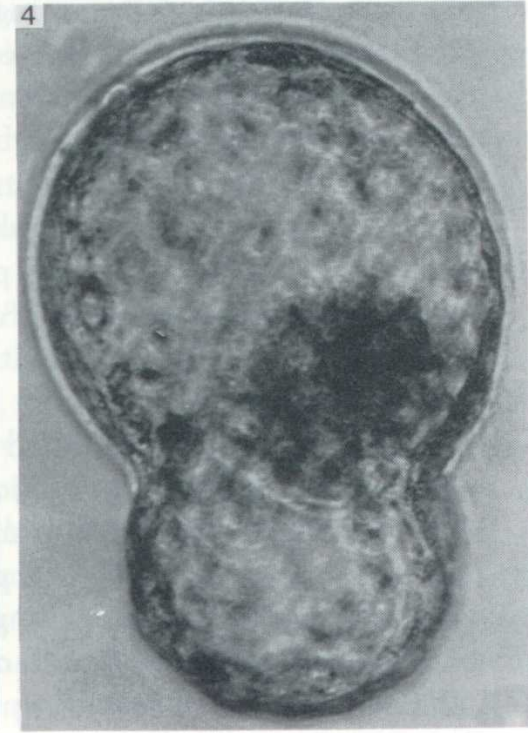
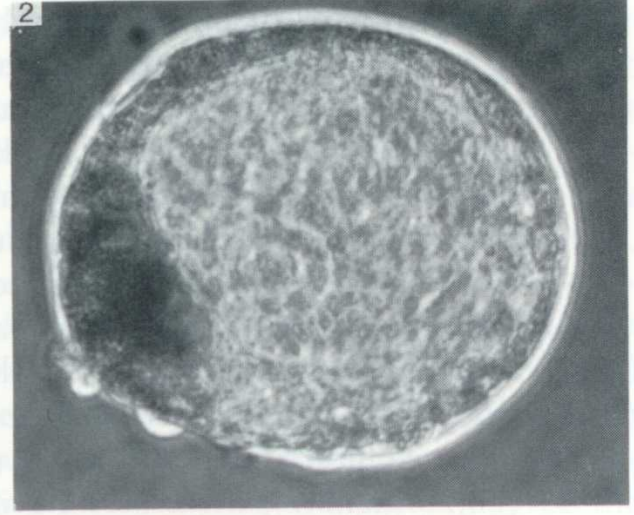
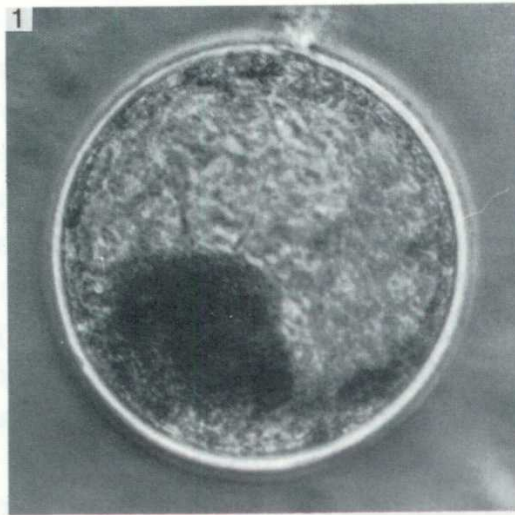


blastocyst

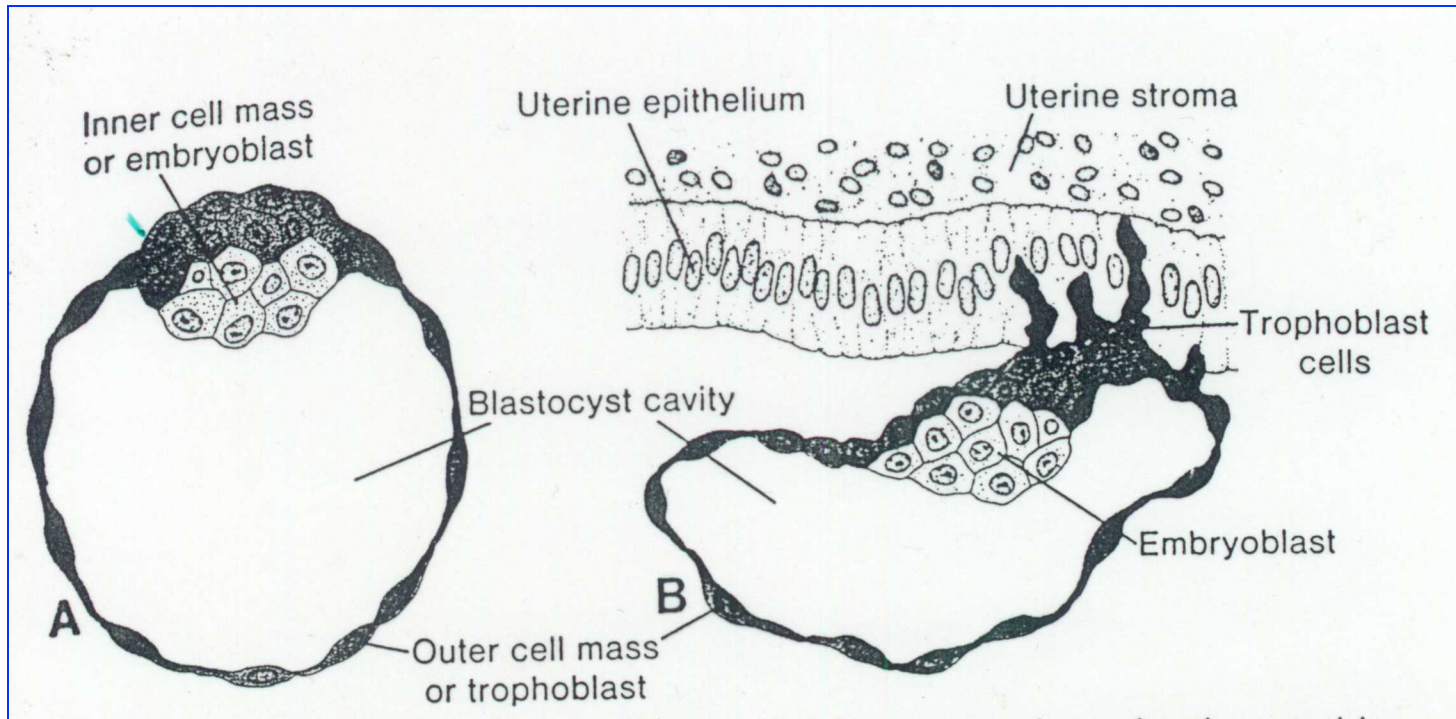
Assessment of bovine embryo quality as a model for human embryo development



The hatching process in a bovine embryo



Embryo implantation to the uterine wall



Times of arrival in the uterine cavity and implantation in different species

Species	Arrival (d)	Implant (d)
Bovine	3-4	12
Bitch	5-8	13-17
Queen	5-8	13-17
Sheep	3	15
Mare	5-6	25-30
Sow	2	2
Woman	4	5-6

Causes of embryo mortality

- Genetic factors:
 - chromosomal abnormalities
 - single gene defects
- Maternal age
- Endocrine factors
- Nutritional factors
- Stress factors
- Internal environmental factors
- External environmental factors

Embryo metabolism

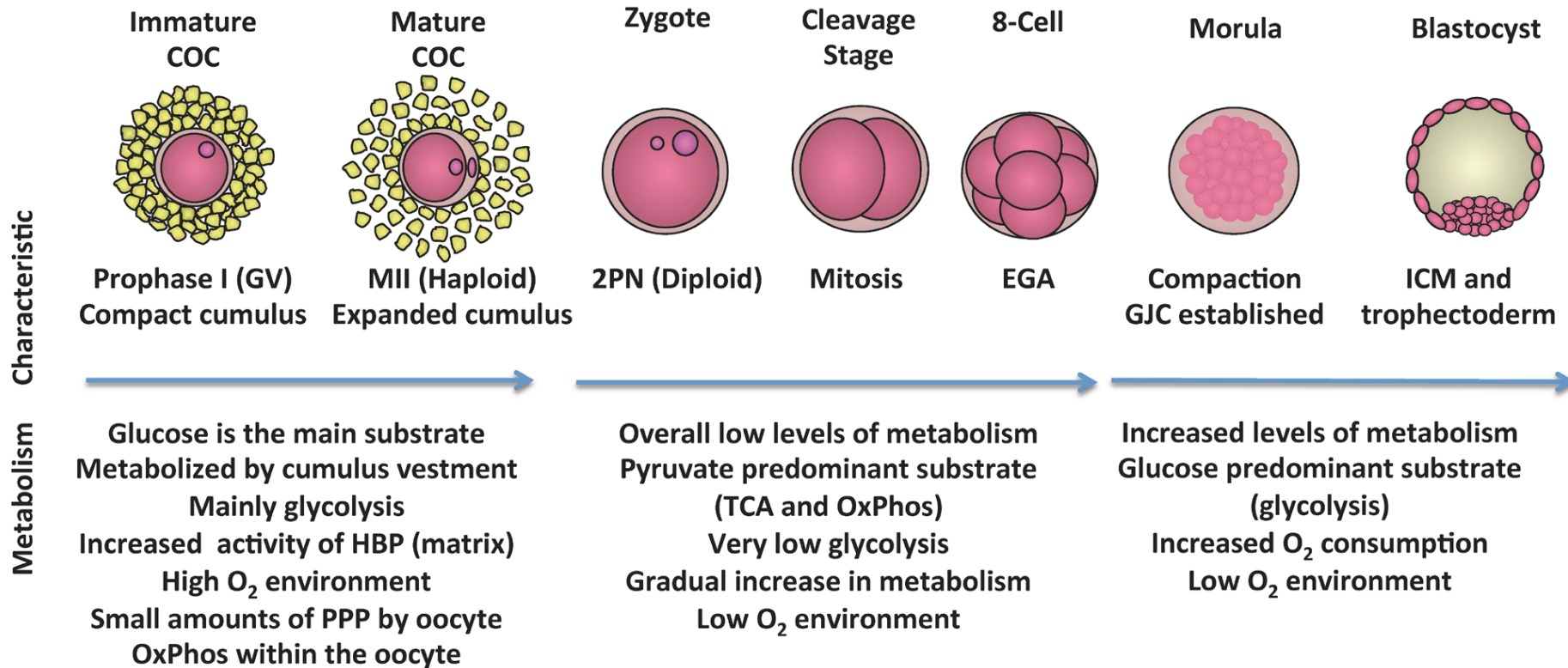
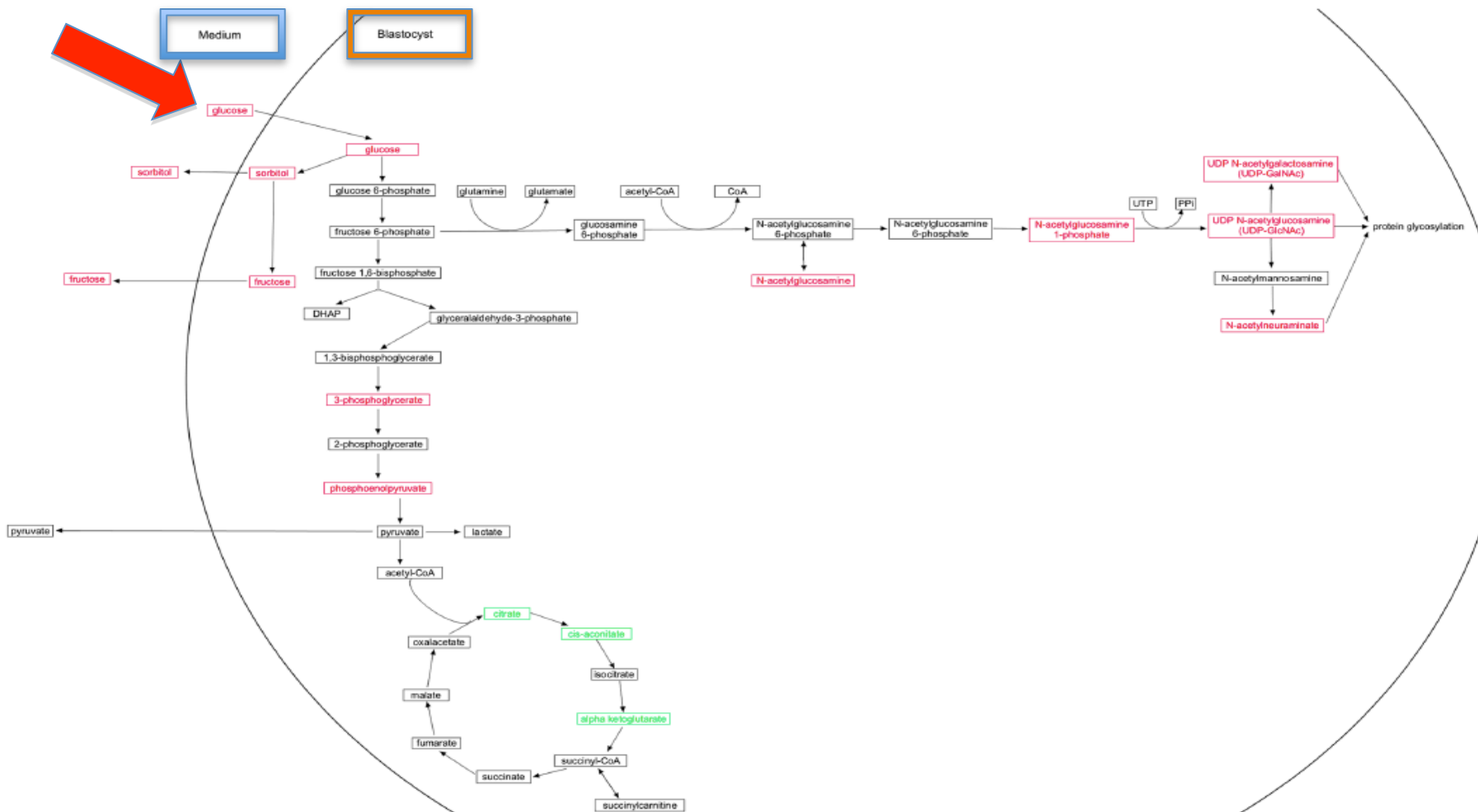


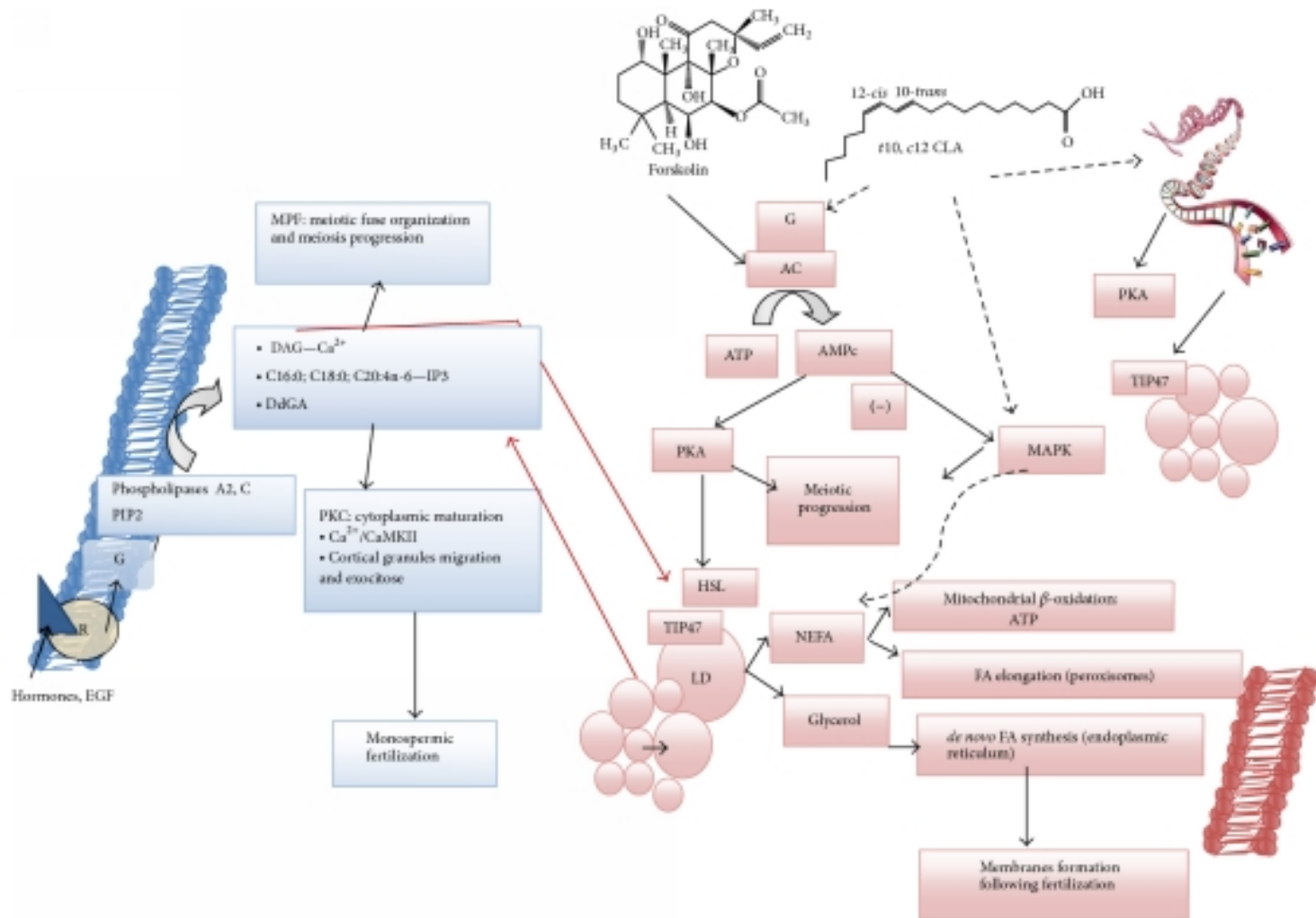
Figure 1. Changes in the metabolism of cumulus oocyte complexes (COCs) and preimplantation embryos. 2PN = 2 pronuclei; GJC = gap junction communication; GV = germinal vesicle; HBP = hexosamine biosynthetic pathway; ICM = inner cell mass; OxPhos = oxidative phosphorylation and TCA cycle = tricarboxylic acid cycle.

Embryo metabolism: the importance of glucose levels



Embryo metabolism:

Fatty acid metabolism (Prates et al., 2014)



Embryo genome activation (EGA) and the developmental arrest

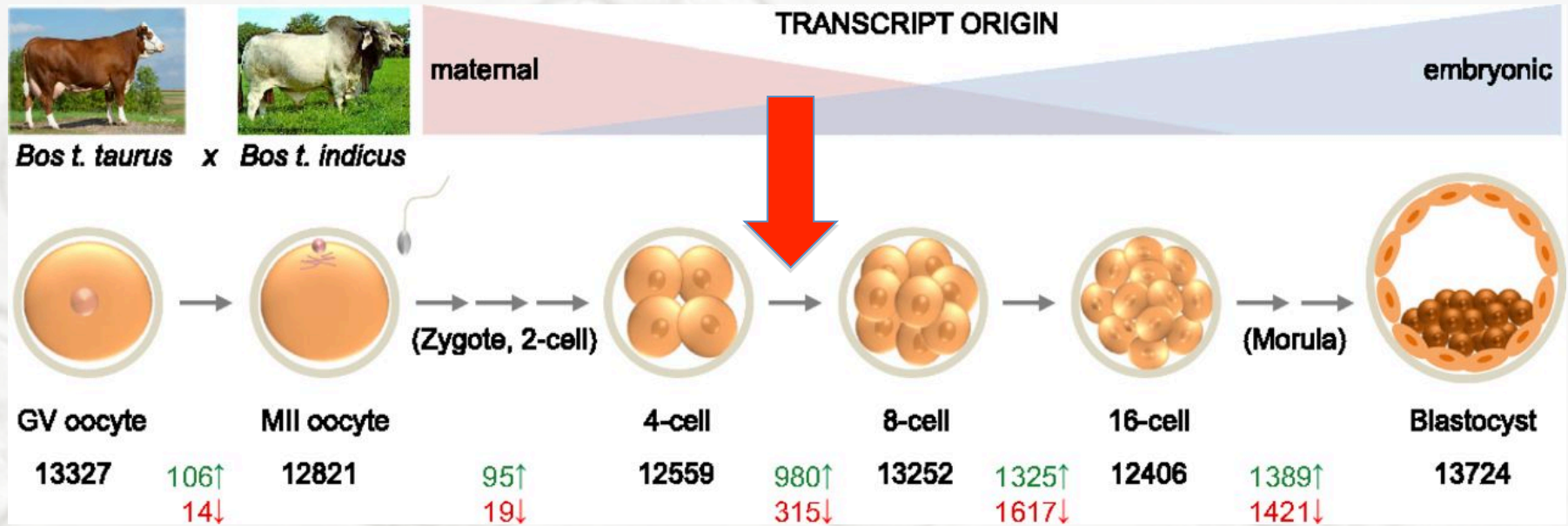
- Embryo genome activation, in different species, occurs at different developmental stages
 - Mouse, Goat, Hamster: 2 cell stage
 - Cow, Sheep: 8-16 cell stage
 - Pig: 4 cell stage
 - Human: 4-8 cell stage

Embryo differentiation

Hierarchical organization of genes coding for factors involved in the differentiation processes:

- **Maternal effects genes** (expressed in the stages comprised from oocyte maturation to initial embryo cleavages)
- **Gap genes** (expressed in subsequent stages and coding for basic cell functions, such as nucleus functions, cytoskeleton elements, membrane channels, ion transporters, surface or secretory proteins, growth factors, receptors, ...)
- **Genes for segmentation and embryo polarity** (genes controlling the formation pattern of the embryo **antero-posterior** axis)
- **Homeotic genes** (homeo-box containing genes, for segment-specific genes)
- **Tissue-specific genes**

Embryo genome activation and developmental arrest



Dean J, NIH Research Images

Fine mapping of genome activation in bovine embryos by RNA sequencing

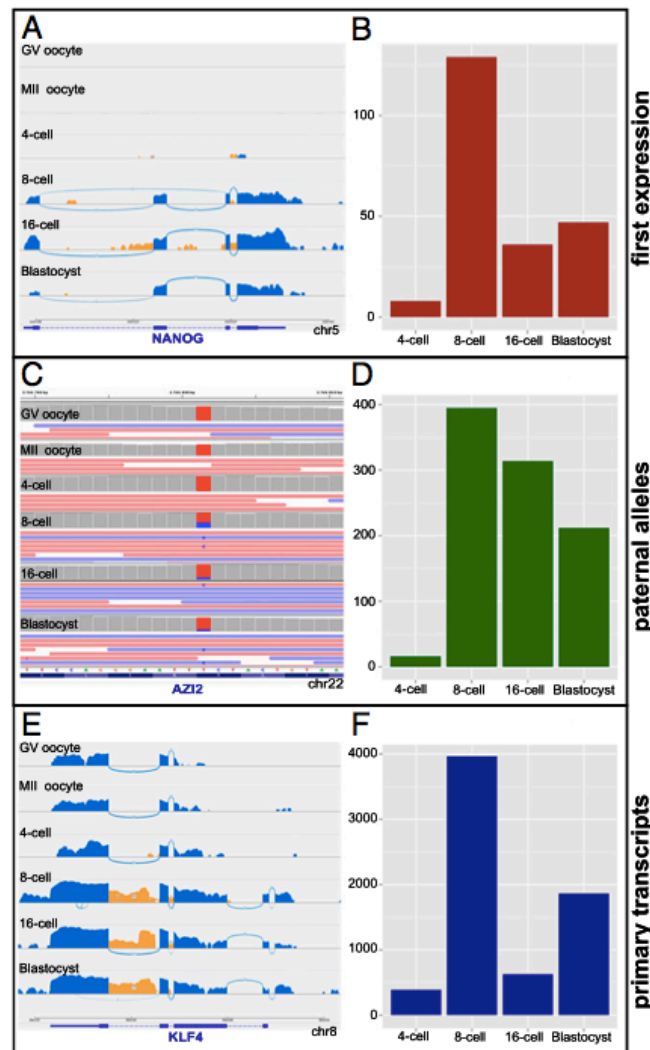
Alexander Graf^a, Stefan Krebs^a, Valeri Zakhartchenko^b, Björn Schwalb^c, Helmut Blum^{a,1,2}, and Eckhard Wolf^{a,b,1,2}

^aLaboratory for Functional Genome Analysis, ^bChair for Molecular Animal Breeding and Biotechnology, and ^cDepartment of Biochemistry, Center for Integrated Protein Science, Gene Center, Ludwig-Maximilians-Universität München, 81377 Munich, Germany

Edited* by George E. Seidel, Colorado State University, Fort Collins, CO, and approved February 7, 2014 (received for review November 18, 2013)

During maternal-to-embryonic transition control of embryonic several hundred transcripts with increased abundance in tran-

Early embryo genome activation and developmental arrest



Fine mapping of genome activation in bovine embryos by RNA sequencing

Alexander Graf¹, Stefan Krebs², Valeri Zakhartchenko³, Björn Schwab⁵, Helmut Blum^{4,1,2}, and Eckhard Wolf^{4,1,2}

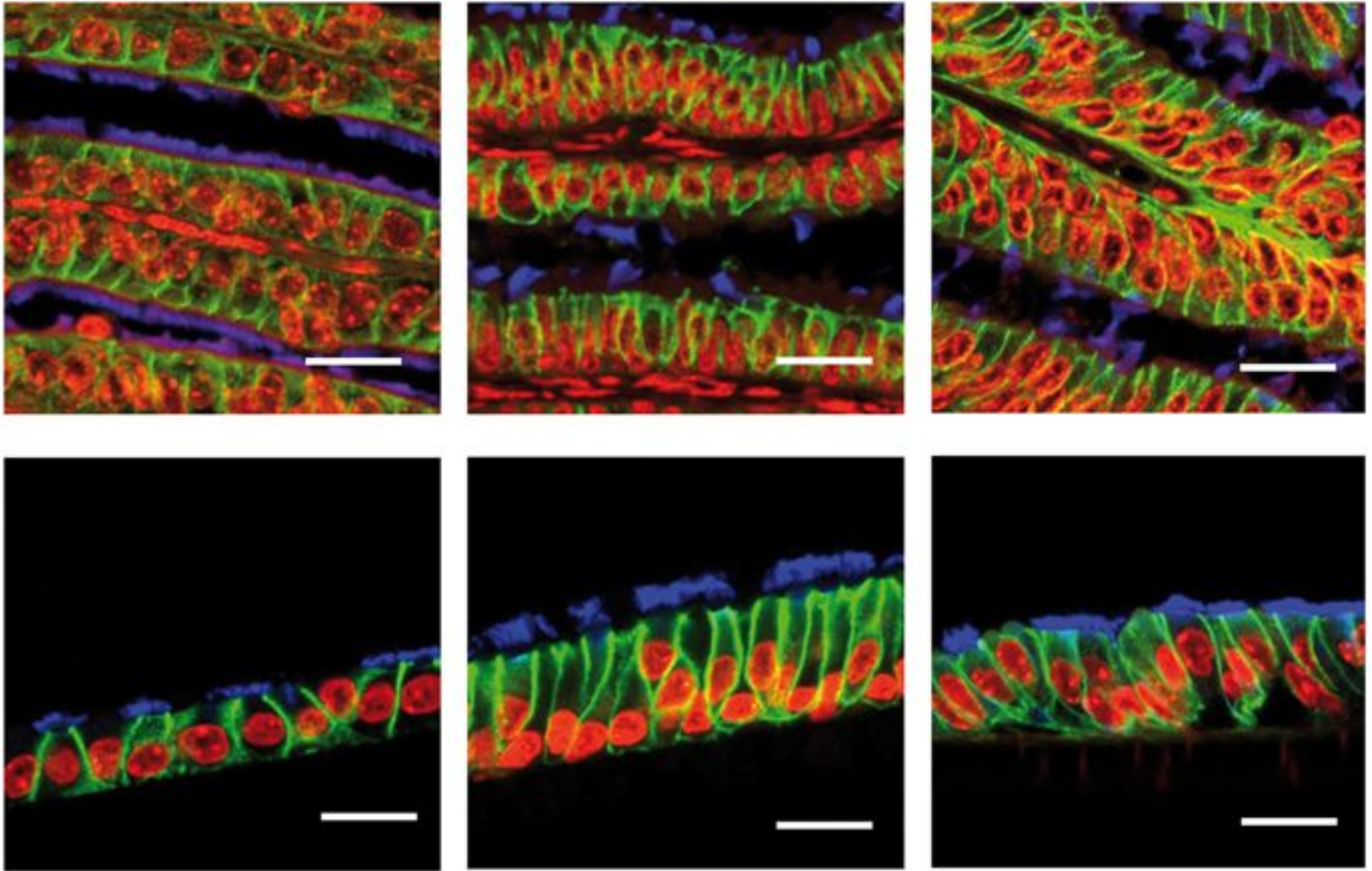
¹Laboratory for Functional Genome Analysis, ²Chair for Molecular Animal Breeding and Biotechnology, and ³Department of Biochemistry, Center for Integrated Protein Science, Gene Center, Ludwig-Maximilians-Universität München, 81377 Munich, Germany

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PNAS

Epithelial markers in murine, porcine and bovine OECs



(Chen et al., 2017)

Embryo quality assessment

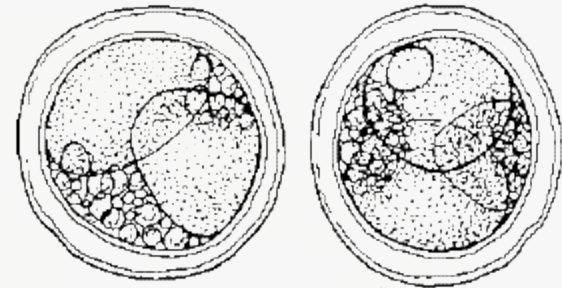
- Morphologic and morphometric analysis
- Correspondence between the culture time and the embryo developmental stage (time course)
- Correspondence between the number of blastomeres and nuclei
- Blastocyst expansion and hatching
- Metabolic tests
- Genomics
- Transcriptomics
- Proteomics
- Metabolomics

Conventional embryo morphology assessment (CMA)



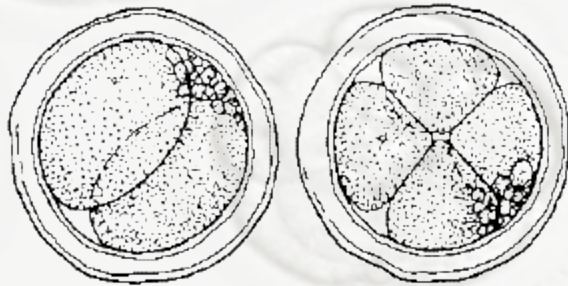
Grade 1

Regular size blastomeres/no fragments



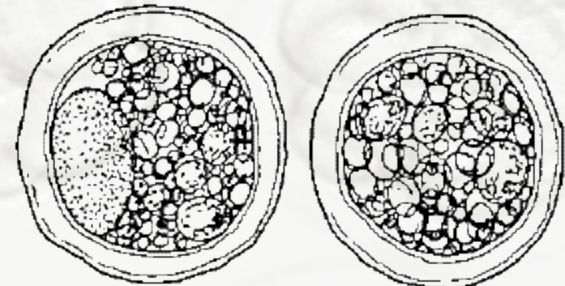
Grade 4

Reg-irreg blastomeres/many fragments



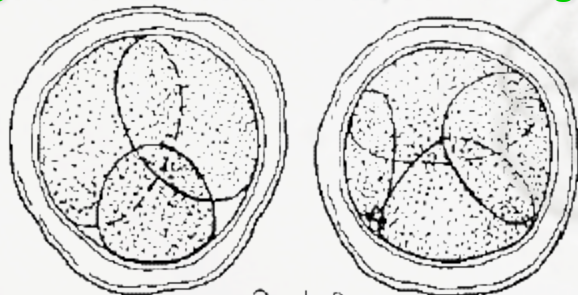
Grade 2

Regular size blastomeres/some fragments



Grade 5

Few blastomeres/severe fragmentation



Grade 3

Irregular size blastomeres

Non invasive pre-implantation genetic screening (NIPGS)

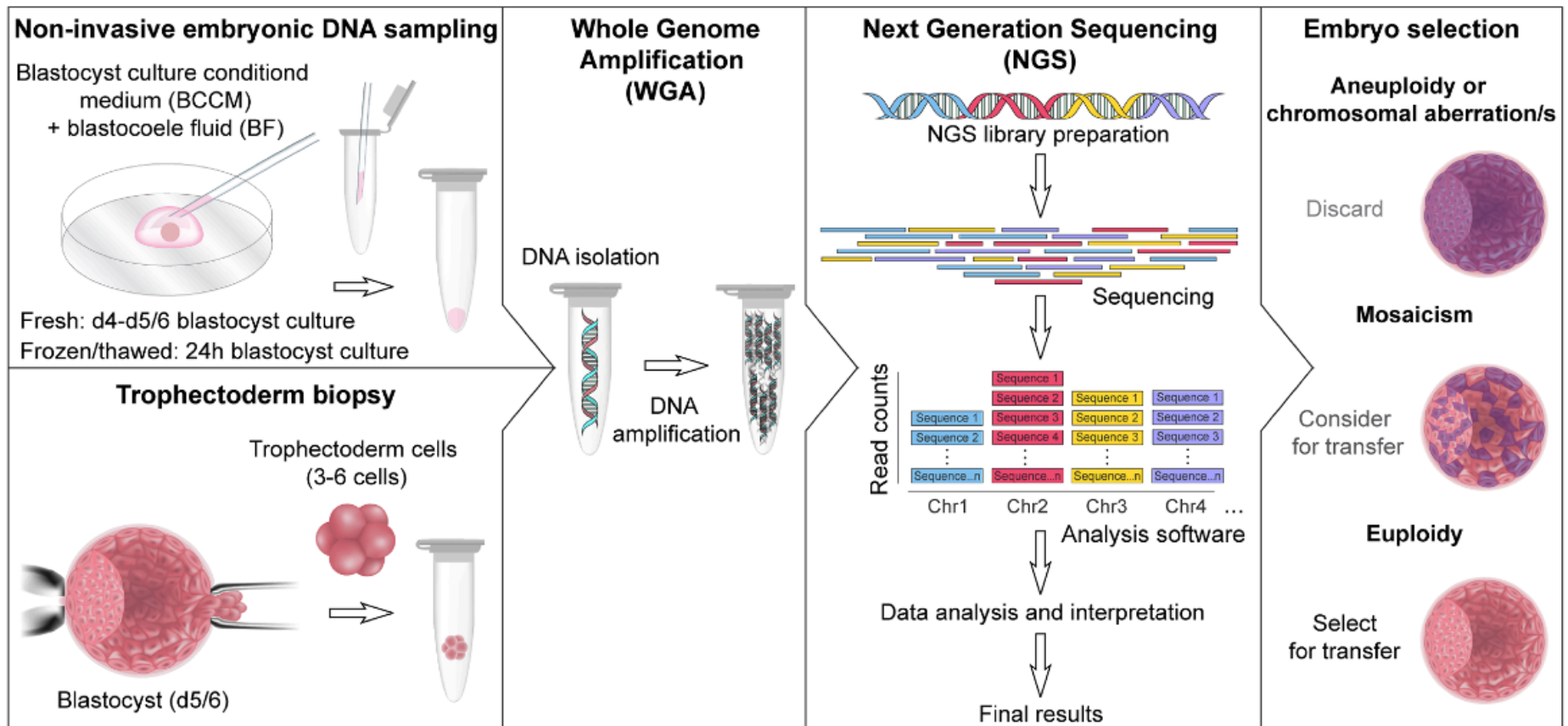


Fig 1. Non-invasive and invasive preimplantation genetic testing workflow.

Metabolic tests to evaluate embryo viability

- **Lactic dehydrogenase (LDH) activity**
- **Glucose, Glutamine, Pyruvate uptake**
- **Oxygen uptake**

References



- “Culturing and evaluating the early embryo” in: Gordon I. Laboratory production of cattle embryo. CABI publications 2003, Chapter 7;
- “Assisted reproductive technologies and embryo culture methods for farm animals” in : Pinkert CA. Transgenic Animal Technology – A laboratory handbook. 2002. Chap 20.